

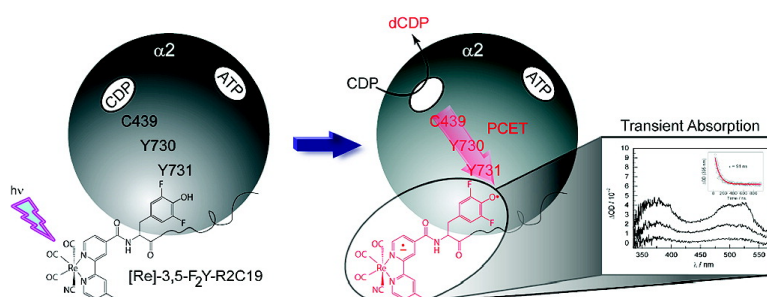
Communication

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Direct Observation of a Transient Tyrosine Radical Competent for Initiating Turnover in a Photochemical Ribonucleotide Reductase

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Proton-coupled electron transfer (PCET) is important to the generation and transport of amino acid radicals within class I *E. coli* ribonucleotide reductase (RNR).^{1–3} RNR catalyzes the conversion of nucleoside diphosphates to deoxynucleoside diphosphates and, as such, plays a crucial role in DNA replication and repair.⁴ The enzyme is composed of two subunits: $\alpha 2$, which contains binding sites for nucleotide substrates and effectors and $\beta 2$, which contains a diiron-tyrosyl radical ($\cdot Y122$) cofactor. A complex between $\alpha 2$ and $\beta 2$ is required for activity and turnover is putatively initiated by the transfer of the oxidizing equivalent from $\cdot Y122$ to a cysteine residue (C439) at the active site in $\alpha 2$, therein generating a transient thiyl radical. A docking model of the individual $\alpha 2$ and $\beta 2$ subunits separate Y122 and C439 by 35 Å;^{5,6} recent PELDOR studies support this long distance.⁷

Several conserved aromatic amino acid residues between Y122 and C439 form $\beta 2(Y122 \leftrightarrow W48 \leftrightarrow Y356) \leftrightarrow (Y731 \leftrightarrow Y730 \leftrightarrow C439)\alpha 2^3$ as a viable pathway for radical hopping in RNR though the precise mechanism defining the electron and proton coupling in radical transport remains to be defined.² To study the PCET of radical transport in RNR, we have engineered photoRNRs. In these constructs, the $\beta 2$ subunit is replaced by its C-terminal, 20-residue peptide (Y-R2C19), which contains both the binding determinant of $\beta 2$ to $\alpha 2$ and the redox active Y356 residue. The RNR is transformed into a photoRNR by appending a photooxidant to Y356 on the peptide. Under these conditions, $\cdot Y356$ formation and RNR turnover may be phototriggered. We have previously used Trp,⁸ benzophenone, and anthraquinone⁹ to phototrigger $\cdot Y356$. These studies show that radical transport in $\alpha 2$ most likely occurs via transfer of both a proton and an electron between the $Y731 \leftrightarrow Y730 \leftrightarrow C439$ residues shown in Figure 1; however, the $\cdot Y356$ radical has not been directly detected. We now improve the photoRNR construct by the use of $Re(bpy)(CO)_3CN$, [Re], as the photochemical $Y\cdot$ generator¹⁰ and the 3,5-difluorotyrosine (3,5-F₂Y) unnatural amino acid¹¹ at “position 356” in the Y-R2C19 peptide shown in Figure 1. With Y-R2C19 modified with [Re], we achieve the highest turnover yet recorded for a photoRNR and we observe, for the first time, a radical, 3,5-F₂Y \cdot , on the peptide bound to $\alpha 2$ that is competent for initiating deoxynucleotide formation.

The [Re]-3,5-F₂Y-R2C19 peptide was prepared using solid-phase peptide synthesis by coupling the N-terminal amino acid to COOH-functionalized bipyridine (bpy) ligand of the [Re] complex (see SI for synthetic details). Nanosecond transient absorption (TA) spectroscopy was employed to investigate photochemical tyrosyl radical generation on the modified peptide. Similar to what has previously been described for the [Re]-3,5-F₂Y model complex,¹² excitation of the full length peptide in aqueous solutions at pH > pK_a(3,5-F₂Y) = 7.2 with 355 nm laser light produces the [Re⁰]-3,5-F₂Y-R2C19 charge-separated state, which decays with a time constant of 70 ns (Figure 2). Of note, the peak at 395 nm resembles that previously observed for 3,5-F₂Y \cdot in dipeptides studies.^{11,12}

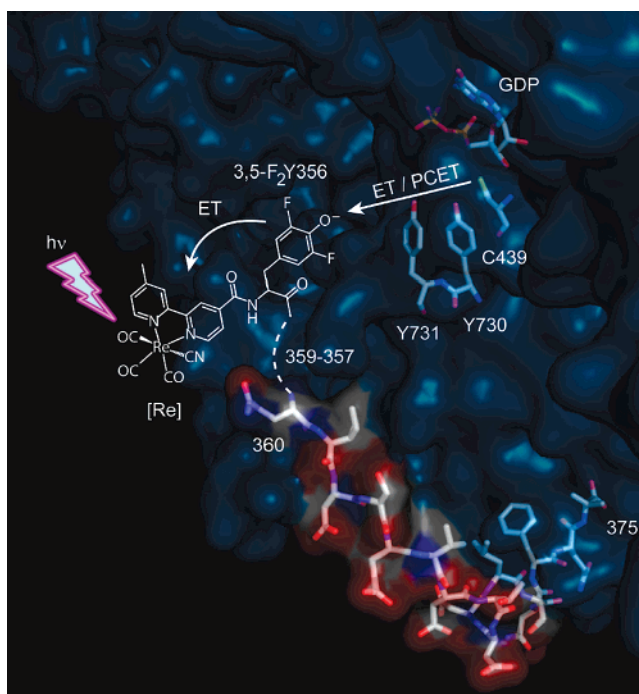


Figure 1. A photoRNR with [Re] as the photochemical radical initiator. Shown is the structure of the $\alpha 2$:Y-R2C19 complex from ref 6. Only peptide residues 360–375 ($\beta 2$ numbering) are visible in the structure. [Re] and 3,5-F₂Y356 have been added for illustrative purposes. Excitation of [Re] with UV light initiates 3,5-F₂Y \cdot formation and radical transport in $\alpha 2$.

The binding of [Re]-Y-R2C19 to $\alpha 2$ was investigated using an established competitive inhibition assay of RNR in the presence of increasing concentrations of peptide.¹³ Figure 3 plots the relative activity of a solution $\alpha 2$ (0.1 μM) and $\beta 2$ (0.2 μM) toward dCDP production as a function of increasing concentrations of [Re]-Y-R2C19. As shown in the figure, this peptide effectively competes with $\beta 2$ for binding to $\alpha 2$. The K_D for the peptide: $\alpha 2$ interaction was estimated as 8 μM using the established competitive inhibition assay to approximate binding.^{9,13} This binding strength is similar to that obtained for other Y-R2C19 peptides, indicating that the large [Re] chromophore does not inhibit binding.

Figure 4 shows the results of the single turnover photochemical RNR assay with [Re]-3,5-F₂Y-R2C19 and [Re]-Phe-R2C19 peptides. A 10:1 peptide/protein ratio was employed to bind the majority of $\alpha 2$ to peptide.¹⁴ Using the estimated K_D of 8 μM and a 1:1 binding model of peptide to α , we calculate that 96% of α in solution is bound to peptide. Experiments were performed by steady-state photolysis of a solutions of 200 μM peptide, 20 μM $\alpha 2$, 3 mM ATP effector, and 0.75 mM [5-³H]-CDP substrate (with 15 mM MgSO₄, 50 mM Tris buffer, and 20% glycerol) using light of $\lambda > 348$ nm. The details of the photochemical assay and data

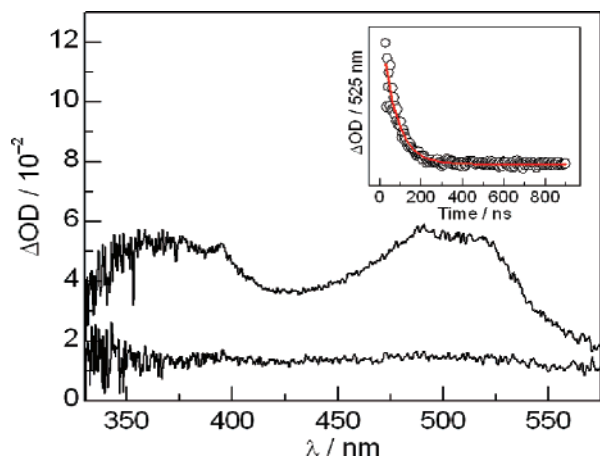


Figure 2. TA spectra of 100 μM [Re]-3,5-F₂Y-R2C19 obtained 65 and 215 ns after a 355 ns flash. Conditions: 50 mM Tris buffer (pH 8.5), 20% glycerol, 1 mM CDP, 3 mM ATP, and 15 mM MgSO₄. The inset shows the single wavelength kinetics traces (○) with single-exponential fit (—) recorded at the wavelength indicated. The peaks at 395 and 525 nm corresponds to 3,5-F₂Y[•] and bpy^{•-} in the charge-separated state.^{10,11}

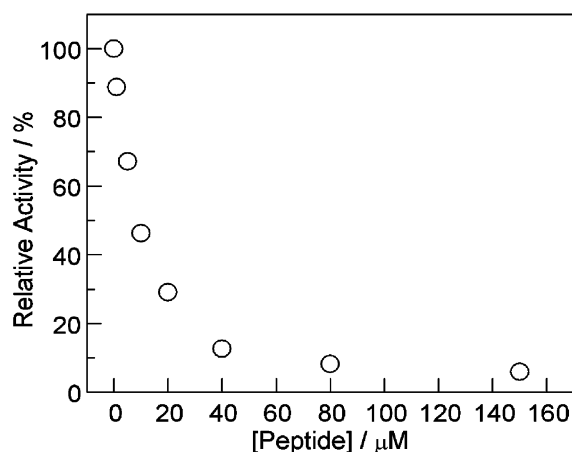


Figure 3. Competitive inhibition binding assay of [Re]-Y-R2C19 to $\alpha 2$. The relative activity of a solution of $\alpha 2$ (0.1 μM) and $\beta 2$ (0.2 μM) was assayed spectrophotometrically as a function of increasing concentration of peptide as described in the Supporting Information.

analysis are provided in the Supporting Information. The addition of glycerol in these experiments served to increase the stability of the protein and resulted in data that was more reproducible.

Photolysis of the [Re]-3,5-F₂Y-R2C19 peptide with $\alpha 2$ resulted in a modest time dependent turnover that approached, after 10 min, 12 and 29% (per α) at pH 8.2 and 7.5, respectively. RNR activity exhibits a pH rate profile with a maximum at pH ~ 8.1 , although the nature of this phenomenon is not understood.¹⁵ Clearly, the photoRNR exhibits a different pH dependence than the wild-type. Mechanistic conclusions regarding the PCET of radical generation and transport cannot be drawn from pH dependent studies of a single photoRNR system, as protein conformations and electrostatics may vary with pH.

The time dependence of the turnover may be a result of the conformational flexibility of the C-terminus of the peptide bound to $\alpha 2$. As shown in Figure 1, this region is disordered and not located in the crystal structure of $\alpha 2$.⁶ The N-terminus of the peptide bound to $\alpha 2$ likely has a high degree of conformational flexibility, and sampling of the conformation that supports radical transport may occur many times during the steady-state photolysis experiment. Similar time-dependent turnover numbers were measured on

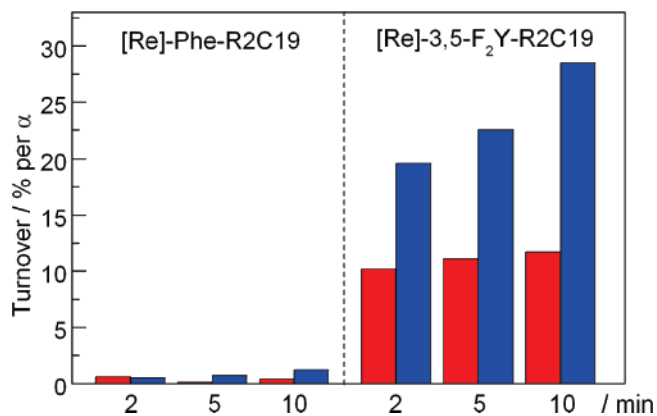


Figure 4. Single turnover assays for $\alpha 2$ with [Re]-(Phe/3,5-F₂Y)-R2C19 peptides in 20% glycerol. Data were collected after 2, 5, and 10 min of photolysis with light of $\lambda > 348$ nm. Red and blue bars are at pH 8.2 and 7.5, respectively.

the minute time scale in the absence of glycerol (data not shown). We therefore do not attribute the time-dependence to an increased viscosity effect.

In contrast, the [Re]-Phe-R2C19 peptide essentially yields no turnover (<1%) throughout the photolysis experiment at both pHs. The lack of turnover with this peptide indicates that the ³[Re]* excited-state cannot directly generate radicals in $\alpha 2$ that are competent for turnover and that radical transport in the [Re]-3,5-F₂Y-R2C19: $\alpha 2$ system must proceed through the transiently generated 3,5-F₂Y[•] radical on the peptide.

The percent turnover obtained with the [Re]-3,5-F₂Y-R2C19 peptide at pH 7.5 after 10 min (29%) is the highest yet recorded for any photoRNR system. Previous photoRNRs were constructed using benzophenone and anthraquinone photooxidants.⁹ Upon excitation, these chromophores were shown to degrade the $\alpha 2$ protein, thus limiting the overall turnover yield. The photochemistry of the ³[Re]* excited state, however, is specific for oxidation of the proximal 3,5-F₂Y residue on the peptide. Furthermore, the [Re]⁰-3,5-F₂Y[•] charge separated state decays via charge recombination on the nanosecond time scale, thereby limiting spurious side reactions of 3,5-F₂Y[•].

Mounting evidence supports that the active $\alpha 2$: $\beta 2$ complex is asymmetric. Studies of disulfide bond formation upon reduction of CDP reveal that the active site cysteines of a single α become oxidized to cystine in the pre-steady state.¹⁶ Pre-steady state and steady state analyses of dCDP formation reveal that that up to 3.0 equiv of dCDP may be formed per $\alpha 2$, rather than the 4 equiv expected for a fully symmetric complex.¹⁷ 3,4-Dihydroxyphenylalanine (DOPA) may be site-specifically incorporated into $\beta 2$ at position 356 and therein serves as a thermodynamic trap during radical transport.^{18,19} The DOPA[•] radical is only observed upon mixing the $\alpha 2$ and $\beta 2$ subunits in the presence of substrate and effector, and only $\sim 50\%$ of $\cdot\text{Y}122$ radical becomes trapped as DOPA[•]. If the active complex of the photoRNR is also asymmetric, we may be limited to a maximum of 50% turnover per α , which is approached here using [Re] as the radical initiator.

Encouraged by the high turnover, we sought to observe the radical upon photolysis of the peptide: $\alpha 2$ complex using TA spectroscopy. Sufficient signal-to-noise ratios were obtained with a minimal amount of spectral averaging (~ 100 spectra) by exciting 100 μM solutions of the peptide with the third harmonic output of a nanosecond Nd:YAG laser (355 nm, 1–2 mJ per pulse). Sample sizes could be limited to 200 μL through the use of a 2×10 mm microcuvette. The white light probe and laser pump beams were

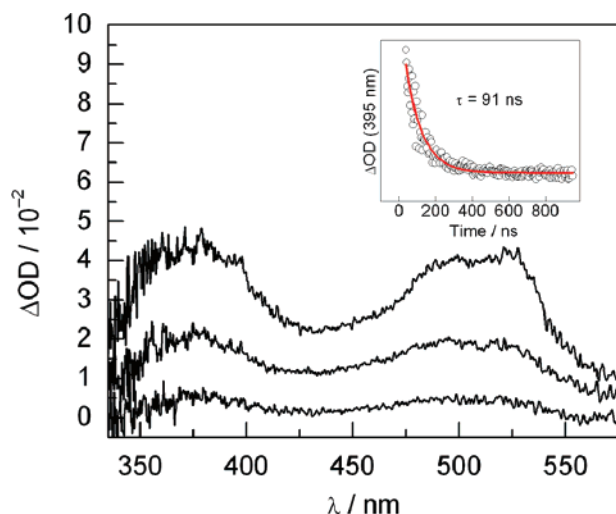


Figure 5. TA spectra of a 100 μM solution of [Re]-3,5-F₂Y-R2C19 in the presence of 140 μM Y731F- α 2, 1 mM CDP, and 3 mM ATP in 20% glycerol at pH 8.2. Spectra were recorded at 65, 115, and 215 ns. The inset is the single wavelength absorbance kinetics recorded at 395 nm.

focused down to 2 mm and strictly overlapped such that they could be passed through the cuvette along the 10 mm path length. This configuration allowed for optimum signal at the low concentrations of the peptide used.

To observe the 3,5-F₂Y• radical on the peptide, bound to α 2, the Y731F- α 2 variant was employed. This mutation inhibits photochemical RNR turnover⁸ by preventing radical transport from •Y356 on the peptide to Y730 in α 2. A 100 μM portion of [Re]-3,5-F₂Y-R2C19 peptide was mixed with 140 μM Y731F- α 2 (with 1 mM CDP, 3 mM ATP, 15 mM MgSO₄, 50 mM Tris, 20% glycerol) at pH 8.2 such that most of the peptide would be bound to protein (87% based on $K_D = 8 \mu\text{M}$). Figure 5 shows the absorption spectra obtained following excitation of this solution with the 355 nm laser pulse. The spectrum at the earliest time point (65 ns) resembles an admixture of the ³[Re]* excited-state and the [Re]⁰-3,5-F₂Y• charge separated state,^{10,12} with a peak at 380 nm (³[Re]*) and a shoulder at 395 nm (3,5-F₂Y•). Single wavelength kinetics recorded at 395 nm could be fit to a single-exponential decay function with a lifetime of 91 ns.

The peak at 395 nm resembles that observed for 3,5-F₂Y• in the TA spectrum of the full length [Re]-3,5-F₂Y⁻-R2C19 peptide free in solution (Figure 2). Under the experimental conditions, 87% of the peptide should be bound. To illustrate that the peak at 395 nm in Figure 5 corresponds to 3,5-F₂Y• bound to α 2, we have calculated a spectrum consisting of 87% ³[Re]* excited-state and 13% [Re]⁰-3,5-F₂Y• (Figure S5), as would be the case if binding of the peptide inhibited radical formation. The peak at 395 nm is hardly visible in this theoretical spectrum, thus the radical observed in Figure 5 must be bound to α 2.

The data of Figure 5 constitute a breakthrough in the investigation of the PCET pathway of radical transport in RNR. The [Re] chromophore, in combination with fluorotyrosines, provides a method for selectively generating tyrosyl radicals on the F_nY-R2C19 peptide bound to α 2 that are competent for turnover and that can be observed with TA spectroscopy. Future experiments will focus on the use of Y730F and C439S- α 2 variants to trap the radical in α 2 and monitor the kinetics for radical hopping from the peptide into the protein. These experiments will ultimately address whether conformational gating or PCET is rate limiting in •Y356 → Y731 radical transport in this system.

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Supporting Information Available: Synthetic details for peptide synthesis/purification and protein isolation; TA studies for the [Re]-(Phe/3,5-F₂Y)-R2C19 peptides; binding assay for [Re]-Y-R2C19 to α 2; details of the photochemical assay. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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- (1) List of Abbreviations: α 2, large subunit of RNR containing substrate and effector binding sites; ATP, adenosine-5'-triphosphate; β 2, small subunit of RNR containing the diiron-tyrosyl radical cofactor; CDP, cytidine-5'-diphosphate; DNA, deoxyribonucleic acid; PCET, proton-coupled electron transfer; PELDOR, pulsed electron-electron double resonance; R2C19, 19-mer, C-terminal peptide tail of β 2; [Re], Re(bpy)(CO)₃CN; RNR, class I *E. coli* ribonucleotide reductase; TA, transient absorption; Tris, tris-(hydroxymethyl)aminomethane.
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